

MCB 5472 Lecture #5: Gene Prediction and Annotation February 24, 2014

Note on the assignment

- Depending on your settings PSI-BLAST can take a while to run
 - Do not leave this until the last minute!
- Recall from Assignment Lecture #1: `nohup` can allow you to leave a job running on the cluster
 - E.g., `nohup [task] & > nohup.out`

Do you have a DNA sequence...

- Limited utility by itself
- Annotations describe what the DNA does
 - Structural: what features are present on the DNA?
 - Functional: what do those features do?

How to annotate: 2 methods

1. From first principles:
 - Experimental data in the literature
 - Algorithmic rules
2. From orthology / homology to previously annotated sequences

Annotation accuracy

- Manual annotation from experimental data in the literature is highly accurate
 - Although not all experiments are unequivocal
- Annotations using algorithms can be quite accurate
 - Depends on the complexity of the problem the algorithm is trying to solve
- Annotations based on orthology relies on the assumption that function is conserved
 - Depends on how rigorously orthologs are defined
 - Depends on functions not changing over time

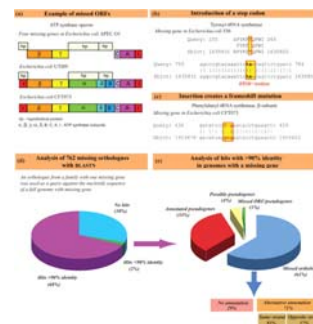
Gene annotation

- Gene and protein annotation is typically algorithmic
- Genes and proteins have specific features that algorithms use to define them
- Algorithms for bacteria and archaea work quite well, eukaryotes more difficult because of additional complexity, e.g., splicing

Prokaryote gene finding

- Glimmer, GeneMark: Markov Models
 - Genes modeled based on differences between coding and non-coding regions
 - E.g., typically start with ATG, end with stop codon
 - E.g., ORF overlap
 - E.g., ribosome binding regions
 - Often have difficulty to decide which strand is coding.
- Prodigal: summed likelihood of finding individual gene features
- Can be challenged by %GC bias
 - Better performance by training on known genome annotations

Fig. 1. Analysis of ORFs missing in one out of 30 completely annotated *Escherichia* genomes.

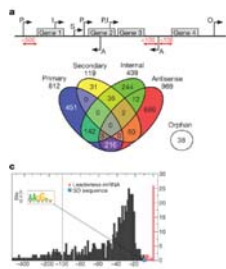


Papadopoulos M S., and Gogarten J P Microbiology
2010;156:1909-1917

Microbiology

Remember: genes are not transcripts!

- 5' mRNA analysis in *Helicobacter pylori* shows much greater transcript diversity than evident from simple gene annotations
- Most NCBI annotations equate genes with transcripts



Sharma et al. 2010 Nature 464:250-255

Eukaryotic gene finding

- E.g., Augustus, GeneMark-ES
- ab initio* methods work less well compared to prokaryotic genes
 - More complicated transcripts (e.g., splice variants)
 - Less information at promoter (e.g., Prodigal uses Shine-Dalgarno sequences; -35 and -10 regions vs. single TATA box)
- NCBI annotations more clearly separate genes (includes pseudogenes), mRNA (typically spliced) & protein (spliced like mRNA)

Adding information to gene annotations

- Combine multiple prediction methods
 - For prokaryotes, typically longest transcript chosen
 - For eukaryotes, typically all splice variants kept
- Search for homologous genes in related taxa
 - True genes will be evolutionarily conserved
 - Annotation errors can be propagated
 - Annotations do not specify the evidence supporting them
- Integrate RNAseq
 - Augustus can incorporate into its predictions directly
 - Rare for prokaryotes
 - Requires genes be expressed and detectable

Metagenomes and single-cell genomes

- Assemblies are typically much more fragmented than those of cultured microbes
- Requires dedicated gene prediction methods
 - Training information often missing/obscured
 - Gene fragments obscure genomic features used for gene prediction

Non-coding RNAs

- Some HMM-based software
 - RNAMMER (ribosomal RNAs)
 - tRNAscan-SE (tRNAs)
- Rfam: database of non-coding RNA families
 - Curated sequence alignments taking into account secondary structures
 - Infernal: software for searching DNA sequence databases using structured RNA molecule profiles
 - Takes RNA secondary structure into account via "covariance models"
 - Sister project to Pfam (see later)

Functional annotations

Manual annotation

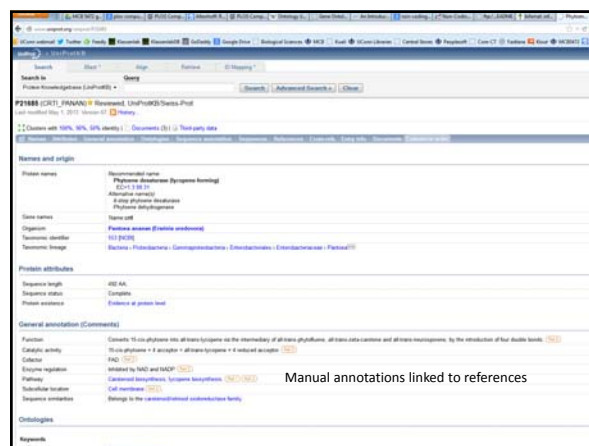
- Low-throughput
- High accuracy

SwissProt

- Started 1986 at the Swiss Institute for Bioinformatics, later developed at the European Bioinformatics Institute
- Goal: providing reliable protein sequences having a high level of annotation
 - Directly curated from literature information
 - Contrast to NCBI: a sequence repository with some automated annotation pipelines
- Current version (2014_02): 542,503 sequences annotated from 22,6190 references

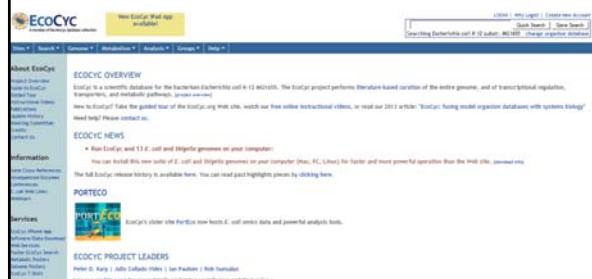
UniProt

- Ultimately manual annotation couldn't keep up, parallel TrEMBL database created using automated annotation
- UniProtKB stores combined SwissProt/TrEMBL databases, incorporates Protein Information Resource (PIR), built on M. Dayhoff's atlas
- Syncs with EMBL/DDBJ/GenBank nucleotide databases
- Hosts several protein annotation schemes
- ExPASy – major proteomics analysis resource
- www.uniprot.org



Ecocyc – an example manually edited model organism database

- <http://ecocyc.org/>

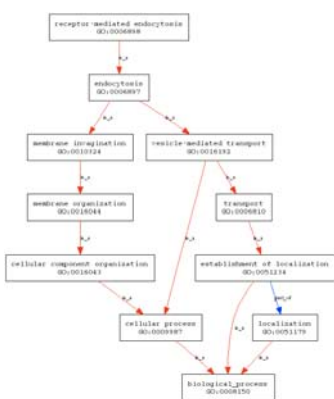


Ontologies

- Manual annotations originally used free-text labels, not standardized
 - Problem: free text is difficult for computers to make use of
- Ontologies: knowledge representation using standardized terms and interrelationships
 - Amenable to computation

E.g., GO

- Controlled vocabulary
- Defined relationships
- “Directed acyclic graph”
 - Links are directional
 - No individually circular paths



<http://topazgen67x09.weebly.com/gene-ontology.html>

Gene Ontology (GO)

- <http://www.geneontology.org/>
- Consortium that defines standardized terms and relationships
- Centered on model organism databases
 - E.g., human, mouse, Drosophila, E.coli
 - Most curation derived from these sources, but do extend more broadly
- Linked and mapped to many other resources
- Used by many computational analysis tools

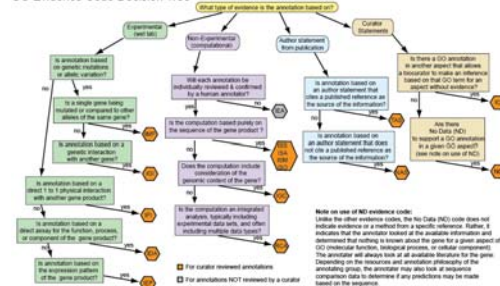
GO domains

- GO is divided into three domains, encompassing three separate functional properties
 - Biological process: what it does
 - Molecular function: how it does it
 - Cellular component: where it does it

GO evidence codes

- GO uniquely has an ontology to describe the evidence supporting annotations

GO Evidence Code Decision Tree



Notes on use of GO evidence codes:
 Unlike the other evidence codes, the GO Data (GO) code does not indicate evidence as a method from a specific reference. Rather, it indicates that the annotation is based on the available information and observation that evidence is required for the gene for a given aspect of its molecular function, biological process, or cellular component. The annotation will always look at all available literature for the gene. Depending on the evidence, the annotation may also look at sequence comparison data to determine if any predictions may be made based on the sequence.

Pfam example

Family: *ABC_tran* (PF00005)

Domain organisation

Below is a listing of the unique domain organisations or architectures in which this domain is found. [More...](#)

Uses

There are 16,039 sequences with the following architecture: *ABC_tran*
 15912: 15912 (Arabidopsis thaliana) Family: ABC transporter proteins (250 residues)
[View alignment](#)

Alignments

There are 16,039 sequences with the following architecture: *ABC_tran*
 15912: 15912 (Arabidopsis thaliana) Family: ABC transporter proteins (250 residues)
[View alignment](#)

Traces

There are 16,039 sequences with the following architecture: *ABC_tran*
 15912: 15912 (Arabidopsis thaliana) Family: ABC transporter proteins (250 residues)
[View alignment](#)

Correlation & model

There are 16,039 sequences with the following architecture: *ABC_tran*
 15912: 15912 (Arabidopsis thaliana) Family: ABC transporter proteins (250 residues)
[View alignment](#)

Species

There are 16,039 sequences with the following architecture: *ABC_tran*
 15912: 15912 (Arabidopsis thaliana) Family: ABC transporter proteins (250 residues)
[View alignment](#)

Structures

There are 16,039 sequences with the following architecture: *ABC_tran*
 15912: 15912 (Arabidopsis thaliana) Family: ABC transporter proteins (250 residues)
[View alignment](#)

Jump to...

[Home](#) [About](#) [Help](#) [Feedback](#)

Pfam example

Family: *ABC_tran* (PF00005)

Curator and family details

The section shows the detailed information about the Pfam family. You can see the definitions of many of the terms in the [glossary](#), and a fuller explanation of the terms used that we use in the [glossary](#) section of the help pages.

Curator

Hand number: 15912
Protein ID: 15912
Accession: E012345678
Number in family: 16,039
Number in full: 16,039
Average length of full alignment: 277
Average identity of full alignment: 43.87 %
Average coverage of the sequence by the domain: 100 %

LMMS information

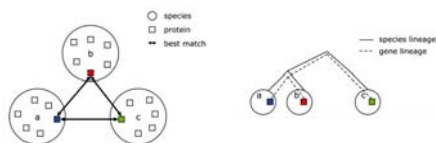
LMMS build command: build method: buildable in (buildable) - build (buildable)
Model details:

Model details	Parameter	Value
Model details	Carbohydrate cut off	23.1
Model details	Truncated cut off	23.1
Model details	Model cut off	23.1

Model length: 277
Family (LMMS) version: 2.1
[Download](#) the raw LMMS for this family

Clusters of Orthologous Groups (COG)

- One of the earliest attempts to define protein families by orthology (Tatusov et al 1997 Science)
- Used BLAST between proteins from multiple genomes to define triangles, i.e., triplets where each is a best match to the others



Kristensen et al. 2010 Bioinformatics 26:1481-1487

COG triangles

- Allows single-direction best hits
- Start with central triangle and add edges whenever possible
- Causes paralogs to be linked
- Allows distant & fast evolving homologs to be linked through intermediates



Solid lines: RBHs
Dotted lines: single direction

Tatusov et al 1997 Science 278: 631-637

COGs

- Bacterial COGs not updated often (last 2003)
- COGs more recently defined for other groups:
 - KOGs (eukaryotes)
 - arCOGs (archaea)
 - POGs (phages)
- Each COG family has a free-text annotation
 - 4873 families total
- Grouped into 24 superfamilies
 - COGs can belong to >1 superfamilies

eggNOG

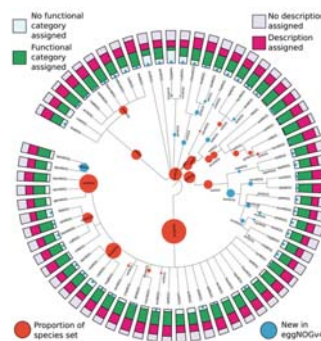
- 'evolutionary genealogy of genes: Non-supervised Orthologous Groups'
- Constructed & maintained by EMBL (Peer Bork)
- Attempt to extend and update COG/KOG database annotations without requiring manual annotations (which do not scale)

eggNOG: method

- Use BLAST/fasta/Smith-Watterman alignments to find best matches
- Represent in-paralogs by single sequences
- Map sequences to COG/KOGs
- Triangle cluster non-matching sequences
- Add single RBH hits to clusters
- Automatically split multi-domain proteins
- Derive annotations by consensus within groups derived from multiple annotation sources

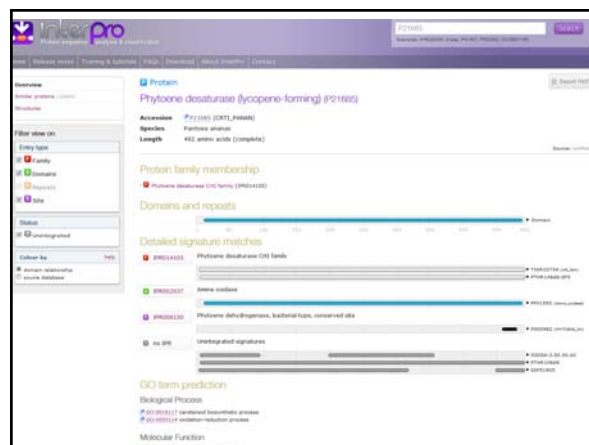
eggNOG:

- http://eggnog.embl.de/version_4.0.beta/
- 107 different annotation levels
- 1.7 million ortholog groups
- 7.7 million proteins
- Probably the currently most comprehensive ortholog database
- Can use to construct PSSMs/HMMs



Interpro

- Classifies proteins according to a combination of multiple protein motifs
- Multiple sources synthesized into single Interpro classification system
 - Four broad annotation types: Family, Domains, Repeats, Sites
- Interpro terms mapped to GO
- InterProScan – resource to annotate proteins using all member databases
 - HMM and regular expression-based classifications



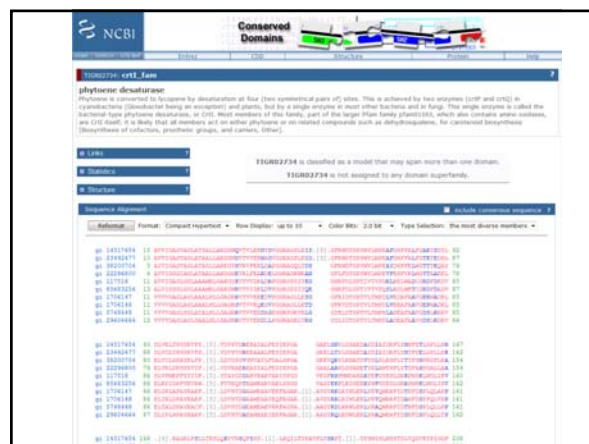
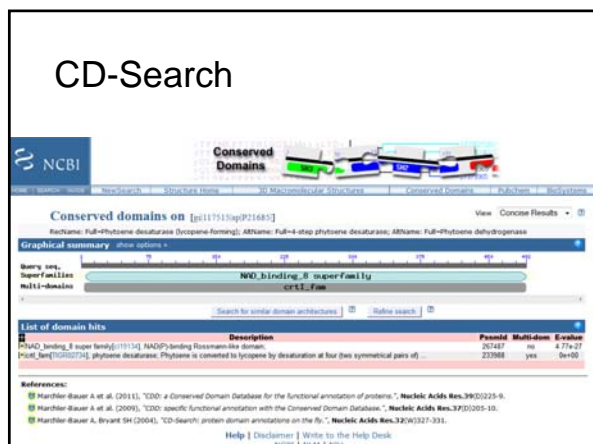
Interpro: member databases

- Pfam (domains, curated; Sanger)
- PROSITE (diagnostic motifs; SIB)
- HAMAP (homologs, curated; SIB)
- PRINTS (conserved motifs; U. Manchester)
- ProDom (domains, automatic via PSI-BLAST; PRABI Villersbanne)
- SMART (domains and architectures esp. signaling, curated; EMBL)
- TIGRFAMs (homologs, curated; JCVI)
- PIRSF (homologs & domains, ; Georgetown)
- SUPERFAMILY (structures, curated, U Bristol)
- CATH-Gene3D (homologs, mapped to structures, automatic via Markov clustering; University College London)
- PANTHER (functional homologs, curated, USC)

Conserved Domains Database (CDD)

- Protein classification database maintained by NCBI
- CDD database based on domains curated by NCBI using structural alignments
- Also includes external resources: Pfam, SMART, COG, PRK, TIGRFAM
- Downloadable PSSMs for each CDD family for querying via RPS-BLAST

CD-Search

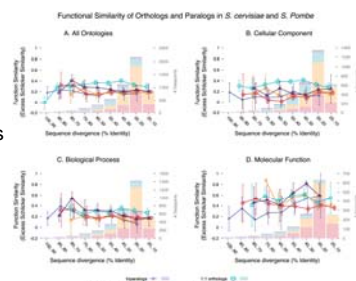


Are functions actually conserved?

- All of the protein annotation methods that we have discussed assume the hypothesis that function is evolutionarily conserved
- But we know that this can be confounded by duplication/loss and xenology
 - Can be addressed by better methods of determining orthology
 - Not typically accommodated by annotation databases
- Even orthologous functions can drift and/or be promiscuous

Are functions actually conserved?

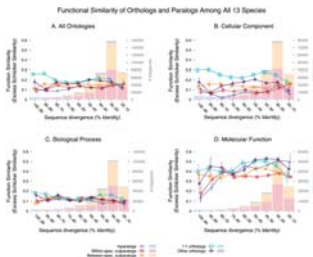
- Compare curated GO annotations of orthologs and paralogs
 - Corrected for annotation biases
- Functions of orthologs more similar than paralogs, but not perfectly



Altenhoff et al. (2012) PLoS Comput. Biol. 8:e1002514

Are functions actually conserved?

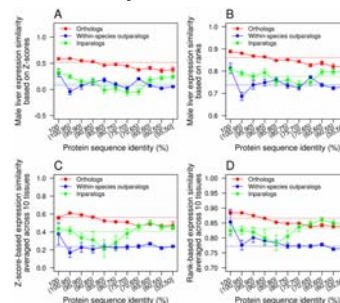
- Same for 13 species instead of just 2
 - Paralogy potentially a greater confounder
- Differences in GO annotation completeness



Altenhoff et al. (2012) PLoS Comput. Biol. 8:e1002514

Are functions actually conserved?

- Define function as expression similarity between same human and mice tissues
- Same trend: ortholog function more conserved than paralogs, not absolute



Chen & Zhang (2012) PLoS Comput. Biol. 8:e1002784



KAAS - KEGG Automatic Annotation Server
An online annotation and pathway mapping

<http://www.genome.jp/kegg/kaas/>

Report

About KAAS

KAAS (KEGG Automatic Annotation Server) provides functional annotation of genes by BLAST comparisons against the manually curated KEGG GENES database. The result contains KEGG (Enzymes) assignments and automatically generated KEGG pathways.

[KEGG Help](#)

Complete or Draft Genome

KAAS works best when a complete set of genes in a genome is known. Prepare query amino acid sequences and use the BDB (3n-directional blast) method to search pathways.

KAAS job request (BDB method)

Partial Genome

KAAS can also be used for a limited number of genes. Prepare query amino acid sequences and use the BDB (single-directional blast) method to search pathways.

KAAS job request (scan method)

KAAS restriction

ETEs

When ETEs are comprehensive enough, a set of conserved motifs can be generated by the [Elassomeria server](#) and used as a gene set for KAAS with the BDB method. Otherwise, use ETEs as file sets with the BDB method.

KAAS job request (BDB method)

KAAS job request (BDB method)

Examples of Results

KEGG assignment

10 Assigned Enzymes

Enzyme: 10 Assigned Enzymes (Detailed description)

Gene name: 10 assigned:

- 10 assigned
- 10 assigned
- 10 assigned
- 10 assigned
- 10 assigned
- 10 assigned
- 10 assigned
- 10 assigned
- 10 assigned
- 10 assigned

KEGG pathway mapping



Reference

Norita, T., Rhee, M., Okada, S., Takahashi, A., and Taniguchi, M.: KAAS: an automatic genome annotation and pathway reconstruction server. *Nucleic Acids Res.* 35, W362-W367 (2007). [[PubMed](#)] [[NA46](#)]

Last updated: Feb 1, 2009

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Biocyc

- Collection of curated metabolic pathways
- Typically smaller modules compared to KEGG
- www.biocyc.org

DATABASE	SCOPE	HIGHLIGHTS	ORGANIZATION
EcoCyc	Escherichia coli K12 (WGS) Metabolic Database	<ul style="list-style-type: none"> Literature curation of complete genome Information from 25,456 publications Transcriptional regulatory network Flux balance metabolic model 	SB International
MetaCyc	Multigenomic Metabolic Pathways and Enzyme Database	<ul style="list-style-type: none"> 2,897 metabolic pathways from 1840 organisms Extensive commentary Information from 37,578 publications 	SB International
HumanCyc	Human specific	<ul style="list-style-type: none"> 271 metabolic pathways 	SB International
AraCyc	Arabidopsis thaliana	<ul style="list-style-type: none"> 400 metabolic pathways Information from 1,530 publications 	S. Rhee, Department of Plant Biology, Carnegie Institution, USA
YeastCyc	Saccharomyces cerevisiae	<ul style="list-style-type: none"> 152 metabolic pathways Information from 1,892 publications 	SIG Genetics, Stanford U., USA
LeidiCyc	Leishmania major parasite	<ul style="list-style-type: none"> 142 metabolic pathways 	BiCi Molecular Science and Biochemistry Institute, University of Melbourne

Biocyc Tier 2: Computationally-Derived Databases Subject to Moderate Curation
 75 databases are available. [see us here](#) [16]

Biocyc Tier 3: Computationally-Derived Databases Subject to No Curation
 2747 databases are available [\(see us here\)](#) and ready for adoption [\(see us here\)](#) by interested scientists for curation and updates

Create Your Own Pathway/Genome Database
 (Research) Curators: [Pathway/Genome Database](#) [see us here](#) [see us here](#)

Metacyc example

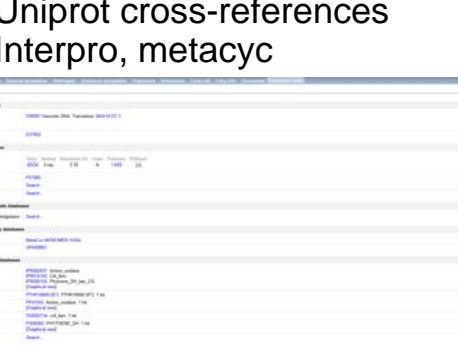
The screenshot displays the Metacyc database interface. At the top, the Metacyc logo and a 'New! Study that too avoided' banner are visible. The navigation bar includes links for 'Home', 'About Metacyc', 'Contact Us', and 'Help'. The main content area is titled 'Metacyc: Enzyme: glucose structure'. It shows the 'glucose' entry under the 'glucose: glycolysis structure' category. The entry details include the enzyme 'glucose 6-phosphate dehydrogenase', the reaction 'glucose + NADP+ -> 6-phosphogluconate + NADPH + H+', and the EC number '1.1.1.49'. The 'References' section lists several scientific papers. The 'Links' section provides links to the 'glucose' entry in the 'KEGG' database and the 'glucose' entry in the 'UniProt' database. The 'Glossary' section defines 'glucose' as a monosaccharide. The 'Structure' section shows a chemical structure of glucose, with the formula $C_6H_{12}O_6$ and the molecular weight '180.156 g/mol'. The 'Properties' section lists the 'glucose' entry in the 'KEGG' database and the 'glucose' entry in the 'UniProt' database. The 'References' section lists several scientific papers. The 'Links' section provides links to the 'glucose' entry in the 'KEGG' database and the 'glucose' entry in the 'UniProt' database. The 'Glossary' section defines 'glucose' as a monosaccharide. The 'Structure' section shows a chemical structure of glucose, with the formula $C_6H_{12}O_6$ and the molecular weight '180.156 g/mol'. The 'Properties' section lists the 'glucose' entry in the 'KEGG' database and the 'glucose' entry in the 'UniProt' database.

Metacyc example

[illegible]

Uniprot cross-references

Interpro, metacyc



The screenshot shows the Uniprot.org website with the following information:

- Uniprot reference:** P04960
- Sequence databases:**
 - EMBL: U00096.1 Genbank: D31472.1
 - CD: 337962
- 3D structure databases:**
 - Protein Data Bank: 1A2A
- Protein families and domains:**
 - InterPro: IPR000001
 - Protein families: IPR000001
 - Protein domains: IPR000001
- Protein families and domains:**
 - InterPro: IPR000001
 - Protein families: IPR000001
 - Protein domains: IPR000001
- Protein families and domains:**
 - InterPro: IPR000001
 - Protein families: IPR000001
 - Protein domains: IPR000001

Annotation process

- Use web to find information about particular proteins
- Use individual tools separately on your genome
 - Allows most customization, proofchecking
 - Standard for eukaryotic genomes
- Use automatic prediction servers
 - Common for prokaryotes
 - E.g., NCBI, IMG (JGI), RAST, Megan, MAGE
 - Each vary slightly in algorithm, user engagement and proofchecking, visualization
- Transfer homology from previously-annotated sequences
 - Can propagate incorrect annotations
 - Can limit coverage